Expression Analysis of miRNAs and Their Potential Role as Biomarkers for Prostate Cancer Detection

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Abstract

Prostate cancer (PCa) is the second most frequent cancer diagnosed in men worldwide. The detection methods for PCa are either unreliable, like prostate-specific antigen (PSA), or extremely invasive, such as in the case of biopsies. Therefore, there is an urgent necessity for reliable and less invasive detection procedures that can differentiate between patients with benign diseases and those with cancer. In this matter, microRNAs (miRNAs) are suggested as potential biomarkers for cancer. MiRNAs have been found to be dysregulated in several different cancers, and these genetic alterations may present specific signatures for a given malignancy. Here, we examined the expression of miR141-3p, miR145-5p, miR146a-5p, and miR148b-3p in human tissue samples of PCa (n = 41) and benign prostatic diseases (BPD) (n = 30) using reverse transcription-quantitative polymerase chain reaction (RT-qPCR). We combined the expression results with patient clinicopathological characteristics in logistic regression models to create accurate PCa predictive models. A model including information of miR148b-3p and patient age showed relevant prediction results (area under the curve [AUC] = 0.818, precision = 0.763, specificity = 0.762, and accuracy = 0.762). A model including all four miRNAs and patient age presented outstanding prediction results (AUC = 0.918, precision = 0.861, specificity = 0.861, and accuracy = 0.857). Our results represent a potential novel procedure based on logistic regression models that utilize miRNA expressions and patient age to assist with PCa diagnosis.

Keywords

expression, miRNAs, biomarkers, predicted model, prostate cancer

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Introduction

Prostate cancer (PCa) is the second most diagnosed cancer in men, only behind lung cancer (Cancer Today, n.d.). Therefore, early detection of PCa is one of the main challenges for this pathology. Prostate-specific antigen (PSA) is a well-accepted biomarker for PCa follow-up, as its expression is highly sensitive to clinical conditions. PSA expression is modified in the presence of several prostate diseases and is not distinguishable between patients with and without a malignant disease, which in turn leads to a high percentage of unnecessary biopsies. Subjecting patients to an invasive procedure may cause bleeding, infections, and, in some extreme case, even death and psychological manifestations such as anxiety and depression (Byun et al., 2022; Cary & Cooperberg, 2013; Hendriks et al., 2017; Minervini et al., 2014; Wu et al., 2021). A robust method for early diagnosis is urgently needed. In this regard, microRNA (miRNA) expression profiling is a promising tool for the detection of various types of cancer due to the ease and reliability of extraction from biological samples, their stability, and measurement accuracy by standard techniques, and this is especially true for PCa, where the necessity for minimally invasive, sensitive, and specific biomarkers is both

Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage). fundamental and pressing (Ambrozkiewicz et al., 2020; Chang et al., 2021; Ignatiadis et al., 2021; Moya et al., 2019; Suer et al., 2019). MiRNAs are small noncoding RNA molecules that regulate the expression of various genes post-transcriptionally by binding to the 3'-untranslated region (UTR) region of target messenger RNAs (mRNAs) (Bartel, 2018).

To identify miRNAs as potential biomarkers, first, it is necessary to identify their specific expression profile in the tissue of patients with and without PCa, thus attributing this deregulation to the carcinogenic events that occur in the prostate and evaluating whether these miRNAs conserved the differential expression in some body fluid and whether this is related to their expression in the specific tissue. In addition, numerous studies have reported the utility of miRNAs for disease detection and prognosis (Byun et al., 2022; Giglio et al., 2021; Kim et al., 2021; Lyu et al., 2019; Porzycki et al., 2018; Suer et al., 2019). miRNAs miR141-3p (Osipov et al., 2016; Xiao et al., 2012), miR145-5p (Wang et al., 2015; Xu et al., 2019), miR146a-5p (Fredsøe et al., 2020; Huang et al., 2017), and miR148b-3p (Feng et al., 2019; Tomeva et al., 2022; Walter et al., 2013) have been suggested as predictive molecules for cancer, and especially in PCa for their role in carcinogenesis, tumor progression, and metastasis. This study analyzes the relative expression of miR141-3p, miR145-5p, miR146a-5p, and miR148b-3p, in tissue samples from patients diagnosed with and without PCa, to assess their potential as biomarkers to distinguish between PCa and benign prostate diseases.

Method

Patient Samples

Prostatic tissue samples were collected from patients with a first presumptive diagnosis of PCa through ultrasoundguided transrectal biopsy and stored in Qiazol (Qiagen) at -20° C for later analysis. Our study consisted of 71

samples: 41 samples from patients with PCa diagnosis (PCa group) and 30 samples from patients diagnosed with benign prostatic disease (BPD group). The BPD group included patients with benign prostatic hyperplasia (n = 7), prostatitis (n = 18), both hyperplasia and prostatitis (n = 4), and intraepithelial neoplasia (n = 1). All patients met the inclusion criteria: Mexican, ≥ 18 years, histopathological diagnosis of PCa or BPD, have not received chemotherapy or radiotherapy, and do not present any other type of cancer. All patients were recruited from the Alvarez & Arrazola Radiologists Clinic in Sinaloa, Mexico, from August 2016 to December 2021. Clinicopathological characteristics such as age, weight, height, body mass index (BMI), PSA, and family history of cancer were collected through direct questionnaires and the clinical database. An expert pathologist provided the diagnosis and Gleason score. All patients were approved by signing an informed consent reviewed and approved by the Ethics and Research Committee of Alvarez & Arrazola Radiologists Clinic (study approval number P-3103).

RNA Extraction and Reverse Transcription (RT) PCR

Tissue samples were used for RNA extraction. Total RNA, including miRNAs, was isolated using the miR-NEasy kit (Qiagen) according to the manufacturer's protocol. The isolated RNA concentration was measured with a GENESYS 10S UV-Vis Spectrophotometer (Thermo ScientificTM). It also allows for evaluating the integrity of the RNA, obtaining information on contaminants, analysis of low or atypical concentrations, and warning of invalid results. The analysis of concentration and integrity was performed in triplicate. RT was performed from 10 ng of total RNA with the TaqMan Advanced miRNA cDNA Synthesis kit (Applied Biosystems) in a T100 Thermal Cycler (Bio-Rad).

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Relative miRNA Expression

The real-time polymerase chain reaction (PCR) technique was carried out using the StepOnePlusTM thermal cycler (Applied Biosystems). For each of the miRNAs, standard curves of five points of serial dilutions 1:10 were made to obtain the efficiencies from the slope. Quantitative estimations for miR141-3p, miR145-5p, miR146a-5p, and miR148b-3p were performed using TaqMan MicroRNA Assays (Applied Biosystems). To select the normalizing miRNA, we used the RefFinder tool, which integrates different algorithms used for the selection of normalizers, identifying miR191-5p as the most appropriate miRNA for our study (Andersen et al., 2004; Pfaffl et al., 2004; Silver et al., 2006; Vandesompele et al., 2002). To obtain relative miRNA expression, we used the method proposed by Taylor et al. (2019) based on the $\Delta\Delta$ Ct method $(2^{-\Delta\Delta Ct} \text{ algorithm})$ (Livak & Schmittgen, 2001). Finally, quantitative analyses using relative expression were performed using log-transformed relative normalized expression.

Statistical Analysis

Student's t test, Mann-Whitney U test, analysis of variance (ANOVA), and Kruskal-Wallis test (when appropriate) were used to compare differences between continuous variables. The chi-square test allowed the identification of relationships between categorical variables. The Pearson and Spearman tests (when appropriate) gave us the correlation coefficient between the following variables: miRNA expression, serum PSA, age, weight, and BMI. All the variables were contrasted among each other to observe relationships. Each miRNA's potential to aid in distinguishing between PCa and BPD groups was investigated using a receiver operating characteristic (ROC) curve analysis and Fagan's nomogram. The Statistical Package for the Social Sciences (SPSS, Inc., Chicago, IL) version 20 software was used for all statistical calculations. Predictive models integrating different variables were carried out through logistic regressions using the orange program, version 3.27.1. Results with a p < .05 were considered statistically significant.

Results

Clinicopathological Characteristics

The average age of PCa patients was 66.55 ± 12.13 years and of the BPD group was 45 ± 6.94 years (p = .008). BMI evaluation yielded an average of 24.94 ± 3.01 kg/m² for PCa patients and an average of 26.56 ± 2.65 kg/m² for BPD patients (p = .399). Concerning the family history of PCa, we found that 14.63% (n = 6) of our PCa patients had at least one family member affected by this



Figure 1. Heatmap Showing Differential miRNA Expression in PCa Samples Compared With BPD Samples. *Note.* PCa = prostate cancer; BPD = benign prostatic diseases.

disease. Regarding the BPD group, 23.33% (n = 7) had relatives with PCa. However, the analysis showed no relation between PCa family history and the presence of PCa (p = .349). PSA analysis showed that the PCa group had an average of $102.92 \pm 283.56 \ \mu g/\mu L$ and the BPD group an average of $41.28 \pm 55.73 \ \mu g/\mu L$ (p = .945). Classifying PCa patients using the five Gleason score groups (International Society of Urological Pathology [ISUP] grade), we observed that 4.88% (n = 2) were classified as 1 (Gleason score ≤ 6), 31.71% (n = 13) as 2 (Gleason score = 3+4), 31.71% (n = 13) as 3 (Gleason score = 4 + 3), 9.76% (n = 4) as 4 (Gleason score = 8), and 21.95% (n = 9) as 5 (Gleason score ≥ 8).

Relative miRNA Expression

The relative expression of miRNAs miR141-3p, miR145-5p, miR146a-5p, and miR 148b-3p was quantified in all samples, and the results are represented in Figure 1. MiR141-3p presented a 2.92 \pm 0.48-fold expression in PCa patients compared with BPD patients; however, no statistical significance was observed (p = .074). When analyzing miR145-5p expression, an expression factor of 0.71 ± 0.15 was identified in PCa patients compared with BPD patients, showing a significant difference (p =.033). In the case of miR146-5p, the PCa patients had underexpression with an expression factor of 0.61 \pm 0.19; however, no significant difference was identified between groups (p = .051). Finally, miR148b-3p was underexpressed in PCa patients with an expression factor of 0.44 \pm 0.28 and a statistically significant difference between groups (p = .001) (Figure 2).

Correlation Analysis

Correlation analyses were performed to establish the relationship between relative miRNA expressions in PCa samples. The analyses did not show a relation between most relative miRNA expressions, as depicted in Figure 3.



Figure 2. Relative miRNA Expression. (A) Comparison of Relative Expression Between PCa and BPD Groups. (B) Percentage of Samples With Underexpression in miR-145-5p and miR-148b-3p. *Note.* PCa = prostate cancer; BPD = benign prostatic diseases; miRNA = microRNA. *p < .05. **p < .01.

However, a correlation was found between miR145-5p and miR146a-5p, with a coefficient of 0.401 (p = .009). Correlations were also performed between relative miRNA expressions and different clinical characteristics of the groups (PSA, weight, age, and BMI); however, no statistical significance was observed (Table 1).

from a prior probability (odds) of 57% (1.3) to a posterior probability (odds) of 90% (9.5). Regarding the negative likelihood ratio, we obtained a value of 0.54, going from a prior probability (odds) of 57% (1.3) to a posterior probability (odds) of 41% (0.7) (Figure 4D). PPV and NPV were 90.81% and 57.72%, respectively.

ROC Curves of Clinicopathological Characteristics

ROC curve analyses were performed to evaluate the capability of different clinicopathological characteristics to discriminate between PCa and BPD. In that regard, PSA had an area under the curve (AUC) of 0.507, which indicates a discrimination capacity of 50.7% (p = .945). Using the Youden index to identify the most appropriate cut-off point, we observed that values of 6.15 g/L have the best sensitivity and specificity (91.7% and 25%, respectively) (Figure 4A). Positive predictive value (PPV) and negative predictive value (NPV) were 62.52% and 68.83%, respectively. The BMI did not show statistical relevance to distinguish the PCa group, obtaining an AUC of 0.403 (Figure 4B).

Performing an ROC curve for age, an AUC of 0.691 was calculated. Results showed that age has a discrimination of 69.1%, which is statistically significant (p = .008) (Figure 4C). Using the Youden index to identify the best cut-off point, it was estimated that from 71 years onward, the sensitivity is 50% and the specificity is 93.1%. Performing the Fagan nomogram with this cut-off point, a positive likelihood ratio of 7.25 was calculated, going

ROC Curves of MiRNAs

ROC analyses were also performed for miR141-3p, miR145-5p, miR146a-5p, and miR148b-3p. Despite promising AUC results, no statistical significances were observed in the case of miR141-3p, miR145-5p, and miR146a-5p (p = .086, p = .068, and p = .070, respectively). However, for miR148b-3p, an AUC of .737 and a p value of .001 were calculated, establishing a discrimination capacity of 73.7% (Figure 5). The most appropriate cut-off point for miR148b-3p was obtained using the Youden index, set at -0.1958, with a sensitivity of 70.7%and a specificity of 70%. When performing the Fagan nomogram, a positive likelihood ratio of 2.36 was observed, going from a prior probability (odds) of 58% (1.4) to a posterior probability (odds) of 76% (3.2). As for the negative likelihood ratio, its value was 0.42, going from a prior probability (odds) of 58% (1.4) to a posterior probability (odds) of 36% (0.6). PPV and NPV were 76.27% and 63.89%, respectively.

Logistic Regression Analysis

Three logistic models were created using the best AUC results to assess the feasibility of distinguishing between



Figure 3. Correlation Analysis Between Different Relative miRNA Expressions in PCa Samples.

Table 1. Correlation between Relative Expression of minings and Clinicopathological Characteristics.							
Variable	Statistics values	miR141-3p	miR145-5p	miR146a-5p	miR148b-3p		
PSA	Correlation coefficient	.008	067	078	.353		
	p-value	.970	.754	.718	.091		
Weight	Correlation coefficient	131	069	.464	038		
	p-value	.641	.800	.070	.889		
Age	Correlation coefficient	069	120	.135	.094		
	þ-value	.690	.471	.419	.574		

-.066

.817

.223

.443

Table I. Correlation Between Relative Expression of miRNAs and Clinicopathological Characteristics

Note. PSA = prostate-specific antigen; BMI = body mass index.

Correlation coefficient

p-value

BMI

PCa and BPD. Model 1 was composed of miR141-3p, miR145-5p, miR146a-5p, miR148b-3p, and age; model 2

was composed of miR141-3p, miR145-5p, miR146a-5p, miR148b-3p, age, and PSA; and model 3 consisted of

.488

.065

-.006

.982



Figure 4. ROC Curve Analysis of Clinical Characteristics: (A) PSA ROC Curve, (B) BMI ROC Curve, (C) Age ROC Curve, and (D) Fagan's Nomogram of Age.

Note. ROC = receiver operating characteristic; PSA = prostate-specific antigen; BMI = body mass index. **p < .01.

miR148b-3p and age. With the assistance of these three models of logistic regression discrimination, we observed that Model 1 had the best ability to distinguish between PCa and BPD, having the highest precision and specificity. In contrast, when incorporating PSA (Model 2), precision and specificity decreased. Importantly, Model 3, composed of only two variables, also yielded an acceptable discrimination capacity (Table 2).

Discussion

We identified that miRNAs miR145-5p and miR148b-3p were differentially expressed between patients with PCa and BPD, suggesting a potential capacity to discriminate between pathologies. When performing analyses to identify the predictive value of these two miRNAs, it was observed that miR148b-3p was the most relevant to distinguish between PCa and benign conditions. Several



Figure 5. ROC Curve Analysis of miR141-3p, miR146a-5p and miR 148b-3p to Differentiate Between PCa and BPD. Note. PCa = prostate cancer; BPD = benign prostatic diseases. **p < .01.

Table 2.	Logistic F	Regression	Parameters	of	Three	Models.
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Model	AUC	Classification accuracy	Precision	Specificity	PPV (%)	NPV (%)
I	0.918	0.857	0.861	0.861	89.42	81.95
2	0.918	0.81	0.823	0.818	85.53	77.04
3	0.818	0.762	0.763	0.762	81.39	70.21

Note. AUC = area under the curve; PPV = positive predictive value; NPV = negative predictive value.

studies had focused on miR148b-3p expression in cancer; however, there are contrasting conclusions attributing it functions either as a tumor suppressor or as an oncogenic miRNA. Shan et al. (2021) observed that this miRNA was overexpressed in exosomes produced by cancer-associated fibroblasts. In lung cancer, miR148b-3p expression was significantly associated with tumor size and tumor grade; in addition, it was observed that patients with a high expression of miR148b-3p were in better health compared with those with a low expression. Thus, miR148b-3p was suggested as a possible prognosis biomarker (Huang et al., 2017). Notably, there are only a few studies involving miR148b-3p and PCa, all of which highlight this miRNA's potential as a biomarker. Walter et al. (2013) observed that miR148b-3p was significantly underexpressed in high-grade prostate tumors (Gleason scales of 4 + 4 and 4 + 5), suggesting its potential in prognosis for this pathology.

In the present study, we observed that miR148b-3p was significantly underexpressed in PCa patients when compared with BPD patients, which is consistent with previous studies (Feng et al., 2019; Tomeva et al., 2022; Walter et al., 2013). In the preceding works, we observed a correlation between miR148b-3p expression and genes PSA and PCA3, which are used as prognosis biomarkers for this disease (Arámbula-Meraz et al., 2020). However, no studies have analyzed its potential as a detection biomarker. Our results demonstrate that this miRNA has potential for PCa detection, yielding an AUC of 0.737 in the ROC curve, with a sensitivity of 70.7%, specificity of 70%, PPV of 76.27%, and NPV of 63.89%. MiR148b-3p is directly involved in mechanisms related to PCa development. Tomeva et al. (2022) observed that this miRNA was significantly underexpressed in tumors with mutations in the androgen receptor (AR). Moreover, Feng et al. (2019) observed that miR148b-3p was underexpressed in tumor tissue; in addition, they identified that it might bind to the 3'-UTR region of KLF4, a transcription factor that regulates gene expression. They also demonstrated that miR148b-3p inhibits the growth of PCa tumor cells in vivo. It is important to mention that in 2016, Siu et al. (2016) observed that KLF4 was overexpressed in PCa cell lines, directly promoting AR overexpression and exhibiting the close relationship between miR148b-3p deregulation and these fundamental mechanisms in the prostate. Although individually miRNAs miR141-5p, miR145-5p, and miR146a-5p were not statistically significant in the predictive analysis, they yielded better results than those obtained with PSA. This agreed with previous reports such as Giglio et al. (2021) and Yang et al. (2022), who obtained AUCs of 0.50 and 0.542 for PSA, respectively, manifesting the importance of exploring new and better biomarkers. By combining our most significant variables in a single predictive model and logistic regression, we obtained a higher AUC (0.918) with better precision and specificity; this model included all four studied miRNAs and patient age. When limited miRNA information is at hand, we proved that simpler predictive models might also be useful. In this matter, our Model 3, which only included miR148b-3p and age as variables, showed relevant specificity and accuracy values (with an AUC of 0.818). These are interesting results because obtaining such high AUC values is not common. Previous and valid works have reported their best miR-NA's AUCs ranging from 0.620 to 0.720 for individual miRNAs and from 0.694 to 0.870 for models that combined multiple variables (Abramovic et al., 2021; Damodaran et al., 2021; Lyu et al., 2019; Suer et al., 2019). These results must be evaluated in other and larger populations to identify that the expression patterns are maintained. In addition, it is necessary to analyze these miRNAs in a body fluid to evaluate their potential to distinguish between benign disease and cancer.

Conclusion

Our results highlight the importance of analyzing miRNAs to distinguish between patients with and without PCa. Mainly, miRNA148 is a potential marker to identify patients with PCa. In addition, multiple markers that include the expression of miRNAs and clinical characteristics show high sensitivity and specificity to select patients with PCa.

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Declaration of Conflicting Interests

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